### Repetitive DNA and next-generation sequencing: computational challenges and solutions

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## Outline

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- De novo genome assembly
- RNA-seq analysis
- Conclusions

### Abstract

- Repetitive DNA are abundant in a broad range of species, from bacteria to mammals, and they cover nearly half of the human genome.
- Repeats have always presented technical challenges for sequence alignment and assembly programs.
- Next-generation sequencing projects, with their short read lengths and high data volumes, have made these challenges more difficult.
- We discuss the computational problems surrounding repeats and describe strategies used by current bioinformatics systems to solve them.

### Repeats

- A repetitive sequence in the genome. (> 50% in human genome)
- Although some repeats appear to be nonfunctional, others have played a part in human evolution, at times creating novel functions, but also acting as independent, 'selfish' sequence elements.
- Arised from a variety of biological mechanisms that result in extra copies of a sequence being produced and inserted into the genome.

Repeat class	Repeat type	Number (hg19)	Cvg	Length (bp)
Minisatellite, microsatellite or satellite	Tandem	426,918	3%	2-100
SINE	Interspersed	1,797,575	15%	100-300
DNA transposon	Interspersed	463,776	3%	200-2,000
LTR retrotransposon	Interspersed	718,125	9%	200-5,000
LINE	Interspersed	1,506,845	21%	500-8,000
rDNA (16S, 18S, 5.8S and 28S)	Tandem	698	0.01%	2,000–43,000
Segmental duplications and other classes	Tandem or interspersed	2,270	0.20%	1,000-100,000



Box 1 | Repetitive DNA in the human genome

## Genome resequencing projects

- Study genetic variation by analysing many genomes from the same or from closely related species.
- After sequencing a sample to deep coverage, it is possible to detect SNPs, copy number variants (CNVs) and other types of sequence variation without the need for de novo assembly.
- A major challenge remains when trying to decide what to do with reads that map to multiple locations (that is, multireads).



Figure 1 | Ambiguities in read mapping.

# Multi-read mapping strategies

- Essentially, an algorithm has three choices for dealing with multi-reads:
  - 1. Ignore them
  - 2. The best match approach (If equally good, then choose one at random or report all of them)
  - 3. Report all alignments up to a maximum number, d (multi-reads that align to > d locations will be discarded)



Figure 2 | Three strategies for mapping multi-reads.

### De novo genome assembly

- Set of reads and attempt to reconstruct a genome as completely as possible without introducing errors.
- NGS vs. Sanger sequencing

	NGS	Sanger			
Length	50~150 bp	800~900 bp			
Depth of coverage	High	Lower			
Hard!					



http://www.data2bio.com/images/assembly\_bg.png

## Problems caused by repeats

- Caused by short length of NGS sequences
  - Repeat length > Read Length

					IN	Hullall. 250~5000p
Repeats				(		
Reads						
	?		?			
		?				?
						NGS: 50~150bp

 If a species has a common repeat of length *N*, then assembly of the genome of that species will be far better if read lengths are longer than *N*.

 $U_{11000}$ , 250 500hn

**N** *I* 

### Problems caused by repeats

- Current Assemblers
  - Overlap-based assembler
  - De Bruijn Graph assembler
- Reads  $\rightarrow$  Graph  $\rightarrow$  Traverse & Reconstruct
- Repeats cause branches → Guess!
  - 1. False Joins
  - 2. Accurate but fragmented assembly. (Short contigs)

#### Ba Assembly graph



**Bb** Correct assembly



Bc Misassembly







#### Cb Correct assembly



Cc Misassembly



Figure 3 | Assembly errors caused by repeats (B, C)

### Problems caused by repeats

- The essential problem with repeats is that an assembler cannot distinguish them.
- The only hint of a problem is found in the paired-end links.



 Recent human genome assemblies were found 16% shorter than the reference genome. The NGS assemblies were lacking 420 Mbp of common repeats.

# Strategies for handing repeats

- 1. Use mate-pair information from reads that were sequenced in pairs.
- 2. The second main strategy: compute statistics on the depth of coverage for each contig
  - Assume that the genome is uniformly covered.



# **RNA-Seq Analysis**

- High-throughput sequencing of the transcriptome provides a detailed picture of the genes that are expressed in a cell.
- Three main computational tasks:
  - Mapping the reads to a reference genome
  - Assembling the reads into full-length or partial transcripts
  - Quantifying the amount of each transcript.

# Splicing

- Spliced alignment is needed for NGS reads.
  - → Aligning a read to two physically separate locations on the genome.
  - For example, if an intron interrupts a read so that only 5 bp of that read span the splice site, then there may be many equally good locations to align the short 5 bp fragment.







## Gene expression

- Gene expression levels can be estimated from the number of reads mappig to each gene.
- For gene families and genes containing repeat elements, multi-reads can introduce errors in estimates of gene expression.



## Conclusions

- Repetitive DNA sequences present major obstacles to accurate analysis in most of sequencing-based experimental data research.
- Prompted by this challenge, algorithm developers have designed a variety of strategies for handling the problems that are caused by repeats.

## Conclusions

- Current algorithms rely heavily on paired-end information to resolve the placement of repeats in the correct genome context.
- All of these strategies will probably rapidly evolve in response to changing sequencing technologies, which are producing ever-greater volumes of data while slowly increasing read lengths.

### Thank you very much.

The end.